- [1] LI-COR Biotechnology web site. http://www.licor.com/bio/. [bib]
- [2] Protocol-Online web site. <u>http://www.protocol-online.org</u>. [<u>bib</u>]
- [3] B. Alberts et al. Molecular biology of the cell. Garland Science, 4th edition, 2002. [bib]
- [4] M. Brenowitz, M. R. Chance, G. Dhavan, and K. Takamoto. Probing the structural dynamics of nucleic acids by quantitative time-resolved and equilibrium hydroxyl radical "footprinting". *Curr Opin Struct Biol*, 12(5):648-653, Nov 2002. [bib]
- [5] P. A. Clarke. RNA-Protein Interaction Protocols, volume 118 of Methods in Molecular Biology, chapter RNA Footprinting and Modification Interference Analysis, pages 73-91. Humana Press, Aug 1999. [bib]
- [6] P. L. Howard, M. C. Chia, S. D. Rizzo, F.-F. Liu, and T. Pawson. Redirecting tyrosine kinase signaling to an apoptotic caspase pathway through chimeric adaptor proteins. *PNAS*, 100(20):11267-11272, 2003. [bib | DOI | .pdf | http]

Signal transduction pathways are typically controlled by protein-protein interactions, which are mediated by specific modular domains. One hypothetical use of such interaction domains is to generate new signaling pathways and networks during eukaryotic evolution, through the joining of distinct binding modules in novel combinations. In this manner, new polypeptides may be formed that make innovative connections among preexisting proteins. Adaptor proteins are specialized signaling molecules composed exclusively of interaction domains, that frequently link activated cell surface receptors to their intracellular targets. Receptor tyrosine kinases (RTKs) recruit adaptors, such as Grb2 and ShcA, that activate signaling pathways involved in growth and survival, whereas death receptors bind adaptors, such as Fadd, that promote apoptosis. To test the ability of interaction domains of Grb2 (Scr homology 2 domain) or ShcA (phosphotyrosine-binding domain) to the death effector domain of Fadd. We find that these chimeric adaptors can reroute mitogenic or transforming RTK signals to induce caspase activation and cell death. These hybrid adaptors can be used to selectively kill oncogenic cells in which RTK activity is deregulated.

- [7] S.-H. L. Kang, K. Vieira, and J. Bungert. Combining chromatin immunoprecipitation and DNA footprinting: a novel method to analyze protein-DNA interactions in vivo. *Nucleic Acids Res*, 30(10):e44, Jun 2002. [<u>bib</u>]
- [8] M. McPike, J. Goodisman, and J. Dabrowiak. Drug-RNA footprinting. *Methods Enzymol*, 340:431-449, 2001. [bib]
- [9] M. P. McPike, J. M. Sullivan, J. Goodisman, and J. C. Dabrowiak. Footprinting, circular dichroism and UV melting studies on neomycin B binding to the packaging region of human immunodeficiency virus type-1 RNA. *Nucleic Acids Res*, 30(13):2825-2831, Aug 2002. [bib]
- [10] V. M. Petrov and J. D. Karam. RNA determinants of translational operator recognition by the DNA polymerases of bacteriophages T4 and RB69. *Nucleic Acids Res*, 30(15):3341-3348, Sep 2002. [bib]

This file was generated by <u>bibtex2html</u> 1.96.